

(FILE 'MEDLINE, CANCERLIT, EMBASE, BIOTECHDS' ENTERED AT 16:46:19 ON 19
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DEL HIS

L1 7839 S NANOCAPSULE# OR NANOPARTICLE# OR NANOSPHERE#
L2 161085 S CATHETER OR NEEDLE INJECTION OR STENT OR MEDICAL DEVICE
L3 22 S L2 AND L1
L4 17 DUP REM L3 (5 DUPLICATES REMOVED)
L5 2038 S MICROPARTICLE
L6 12 S L5 AND L2
L7 8 DUP REM L6 (4 DUPLICATES REMOVED)

=>

Day : Friday
Date: 12/19/2003
Time: 17:08:38

PALM INTRANET

Inventor Information for 09/845080

Inventor Name	City	State/Country
NAIMARK, WENDY	CAMBRIDGE	MASSACHUSETTS
PALASIS, MARIA	WELLSLEY	MASSACHUSETTS

Appln Info

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Petition Info

Atty/Agent Info

Continuity Data

Foreign Data

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L8: Entry 4 of 15

File: PGPB

May 1, 2003

DOCUMENT-IDENTIFIER: US 20030082161 A1

TITLE: Gene preparations

Detail Description Paragraph:

[0030] A liposome vector may be any vector that can be used as an ordinary liposome vector in gene therapy and includes, for example, a liposome vector obtained by mixing DOTMA, DOPE, DOGS, etc. When a cationic liposome vector is used, transfer efficiency into cells is high. Examples of the fusogenic liposome vector in which a virus and a liposome are fused include a fusogenic liposome vector in which a Sendai virus (HVJ: hemagglutinating virus of Japan) and a liposome are fused, and the like. A gene vector can be obtained by enclosing a gene for transfer into a liposome vector or a fusogenic liposome vector according to a conventional method, for example, described in the above references. The enclosed gene may be in any form that can express the gene in living body and is preferably a form stable in a living body such as a plasmid, etc.

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L8: Entry 4 of 15

File: PGPB

May 1, 2003

PGPUB-DOCUMENT-NUMBER: 20030082161
PGPUB-FILING-TYPE: new
DOCUMENT-IDENTIFIER: US 20030082161 A1

TITLE: Gene preparations

PUBLICATION-DATE: May 1, 2003

US-CL-CURRENT: 424/93.21; 424/491, 435/320.1, 435/455

APPL-NO: 10/ 261618 [PALM]
DATE FILED: October 2, 2002

RELATED-US-APPL-DATA:

Application 10/261618 is a continuation-in-part-of US application 08/981552, filed February 4, 1998, ABANDONED

RELATED APPLICATIONS

[0001] The present application is a Continuation-In-Part of co-pending U.S. application Ser. No. 08/981,552, filed Feb. 4, 1998. U.S. application Ser. No. 08/981,552 is the national phase under 35 U.S.C. .sctn.371 of the prior PCT International application no. PCT/JP96/01824, which has an international filing date of Jul. 2, 1996, which designated the United States of America. The entire contents of all of the above applications are hereby incorporated by reference

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L8: Entry 7 of 15

File: PGPB

May 30, 2002

DOCUMENT-IDENTIFIER: US 20020064520 A1

TITLE: TARGETED ARTIFICIAL GENE DELIVERY

Detail Description Paragraph:

[0092] This experiment demonstrates that a gene-delivery particle such as a virion can be re-enveloped by a new functional surface. UV inactivated Sendai virus was used to provide such a functional retargeting and fusogenic enveloping surface to deliver the genetic content of MoMuLV particle to a novel host. Sendai virus previously has been demonstrated to fuse with bare membrane surfaces, and has been applied as a component of fusogenic liposomes in gene delivery (reviewed in Nakanishi et al.; Journal of Controlled Release 54:61-68 (1998)). UV-inactivated Sendai virus was used to fuse with MoMuLV encoding a green fluorescent protein (GFP) marker gene. When MoMuLV was added to a culture of Hela cells no GFP expression was detectable. However, fusion of MoMuLV with Sendai allowed GFP expression in human Hela cells (>50 cfu/ml), demonstrating transduction of a human cell line by the murine virus MoMuLV. Accordingly, these

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L8: Entry 15 of 15

File: USPT

May 20, 1997

DOCUMENT-IDENTIFIER: US 5631237 A

TITLE: Method for producing in vivo delivery of therapeutic agents via liposomes

Detailed Description Text (14):

The term "HVJ-liposome" refers to a vesicle which has been treated with, or associated with viral coat proteins from the Hemagglutinating Virus of Japan (HVJ) Z strain (Sendai virus; paramyxovirus) using attenuated virus, mutated virus, viral envelopes, viral spikes, viral envelope proteins, viral peptides, viral peptide fragments, recombinant viral proteins, recombinant viral fusion proteins and any combination thereof. Two glycoproteins of Sendai virus, such as HVJ, have been identified and purified. These glycoproteins consist of hemagglutinin-neuraminidase (HN) and fusion (F) proteins and they have been shown to play an important role in viral infection and membrane fusion. The HN protein has hemagglutination and neuraminidase activities and is responsible for absorption of virus to the receptor on host cells. The full-length cDNA for HN has been cloned and sequenced by Miura, et al., 1985, FEBS Letter 188:112-116. The F protein of Sendai virus consists of disulfide linked polypeptides, F.sub.1 and F.sub.2 which are derived by proteolytic cleavage of an inactive precursor, F.sub.0. The F protein cDNA has been cloned and sequenced by Miura, et al., 1985, Gene 38:271-274. HVJ particles from mammalian cell cultures are not fusogenic or infective unless they are subjected to mild trypsin treatment to cleave the F protein to its active form. Cleavage also occurs in the chorioallantoic fluid of egg-passaged HVJ. This cleavage of F.sub.0 into F.sub.1 and F.sub.2 is essential to the fusion activity of HVJ (Uchida, et al., 1988, Exp. Cell Res. 178:1-17). Published work reconstituting fusion liposomes using F and HN glycoproteins has indicated that a ratio of 2:1 (F:HN) results in optimal fusing activity (Nakanishi, et al., 1982, Exp. Cell Res. 141:95-101). In a most preferred embodiment of the present invention, the HVJ proteins are derived from the egg adapted Z strain of HVJ deposited with the ATCC and given accession no. ATCC VA 2388. Those skilled in the art will recognize that other proteins which perform a similar function as those from the HVJ (2) strain ATCC VR 2388, are useful in methods of the present invention; such similarly functioning elements from any virus, bacteria or cell that can mediate membrane fusion, may be substituted for the HVJ-derived elements.